

Urine-based bladder cancer diagnostic: Oncuria™



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Abstract

Bladder cancer is a major healthcare issue in the US. For the avoidance of invasive procedures, naturally voided urine is the most clinically relevant biospecimen for the discrimination of patients with bladder cancer. Voided urine cytology, the gold standard urine-based assay for the detection of bladder cancer, has been used since the 1940's with little modification, despite significant limitations in the clinic. Oncuria™, a multiplex immunoassay for the detection of a bladder cancer protein signature, is a validated assay for the detection of bladder cancer from a voided urine sample. Here, we provide information supporting the development of Oncuria™ in the clinic to assist with the non-invasive identification of bladder cancer patients.

*Discovering and Developing Breakthrough Diagnostics for
Human Diseases*



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Bladder cancer

Epidemiology

Bladder cancer is among the most commonly diagnosed cancers worldwide [1]. Urothelial carcinomas are the most common bladder tumors in Western countries, constituting approximately 95% of all cases [2]. Risk factors associated with the development of urothelial carcinomas are mainly environmental exposure to known carcinogens in tobacco smoke [3], and to chemical compounds in the color and rubber industries [4]. Urothelial carcinoma has a 5-times higher prevalence among men than women and has a 2-times higher prevalence in Caucasians than other racial groups [2,5]. In the US, an estimated 83,730 newly diagnosed cases of bladder cancer and 17,200 deaths from bladder were reported in 2021 [5]. Alarming, both the absolute numbers of cases and deaths from bladder cancer have increased by 45 and 35%, respectively, since 2002 [6]. Moreover, bladder cancer has one of the highest recurrence rates of any tumor type. Due to the prolonged natural history of bladder cancer, it is estimated that >700,000 Americans are burdened with bladder cancer each year. Bladder cancer has the highest lifetime diagnostic and treatment cost of all cancers [7], with estimated expenditures of approximately \$187,000 per case and an annual cost of approximately \$4 billion to the healthcare system [7,8]

Treatment

When diagnosed early as a non-muscle invasive bladder cancer (NMIBC, e.g., Ta/T1/Tis stage lesion), cure by transurethral resection (TUR) with immediate post-operative intravesical chemotherapy instillation is possible in a high percentage of cases: 5-year survival rate >94%. Guidelines for NMIBC with adverse features (number of tumors, tumor size, prior recurrence rate, clinical T stage, presence of carcinoma in situ, and grade) recommend adjuvant intravesical instillation of Bacillus Calmette-Guerin (BCG), a live attenuated tuberculosis vaccine that acts as a non-specific immune system stimulant, which has proven to assist in the eradication of residual disease, reducing recurrence rates and decreasing the progression to muscle-invasive lesions [9,10]. However, despite intravesical treatment, more than half of patients fail to respond and 20% experience disease progression to muscle invasive bladder cancer (MIBC) within a 5-year period [11,12]. Failure to intervene with definitive radical cystectomy prior to progression to MIBC is associated with a significant reduction in long-term survival [13,14]. Patients with MIBC (>T2 lesion) treated with either radical cystectomy or radiation therapy have 5-year survival rates of approximately 50% [15,16]. The field of bladder cancer therapeutics has advanced recently with the approval of immune checkpoint inhibitors and reagents that target specific signaling molecules [17], but the early detection and monitoring of disease state remain paramount.

Molecular pathogenesis of bladder cancer

A broad range of research efforts has provided insight into the genetic alterations that lead to the development of bladder cancer [18], and recent studies from US and European consortia have begun to classify bladder cancer into molecular subtypes based on gene expression profiles [19]. Comparisons across independent cohorts have begun to reveal the underlying biology associated with various classification systems, but significant biologic subgroup heterogeneity remains, and more work is needed before a unified classification system can gain wide acceptance.

Current urine-based diagnostics

Table 1 lists the urine-based assays available for the detection of bladder cancer. There are advantages to using urine as a biological sample for biomarker discovery and detection. Tumors in the bladder wall are continuously bathed in urine, facilitating the release of protein, DNA and RNA, as well as intact exfoliated urothelial cells. Urinalysis also has the advantage of easy repeat sampling. This enables the collection of reliable replicate samples (which cannot be achieved with excised tissue) and allows for temporal sampling which may be useful for investigating disease status post-treatment or at recurrence. A brief description of currently available urinary assays for bladder cancer detection follows.

Voided urinary cytology

The most widely used urine-based analysis for the non-invasive detection of bladder cancer is voided urine cytology (VUC), first reported in 1945 [20]. Since its inception, VUC has changed very little. VUC can be a challenging test to perform since it is dependent on the skills and experience of highly trained cytopathologist. The sensitivity reported for VUC is 30-92% with an accompanying specificity of 93-97% [21-25]. The lack of sensitivity achieved by voided urine cytology is skewed by low-grade and early stage tumors, because these

lesions tend to shed fewer cancer cells into the urine for analysis. It is widely known in the urology field that VUC have insufficient predictive power to be applied to the management of individual patients. Because of these substantial limitations, only approximately 10% of patients presenting with hematuria (blood in the urine) are currently evaluated with VUC [26].

Table 1 Performance of current FDA approved/cleared urine-based assays for the non-invasive detection of bladder cancer [24,25]

Urine Test	Analyte(s)	Sensitivity (%)	Specificity (%)	Assay Platform	Quantitative	Multi-sample capability
Cytology	None	30-92	93-97	Microscopy	No	No
ImmunoCyt	Mucin, CEA	32-100	33-91	Microscopy	No	No
BTA TRAK	Complement factor H	52-78	63-86	ELISA	Yes	Yes
BTA stat	Complement factor H	32-100	63-92	Point of Care	No	N/A
NMP22 Test	NMP22	33-100	41-92	ELISA	Yes	Yes
BladderChek	NMP22	38-86	78-96	Point of Care	No	N/A
UroVysion	Aneuploidy 3,7, 17 and loss of 9p21	51-100	55-100	Complex FISH	No / Yes	No
Oncuria™	ANG, A1AT, APOE, CA9, IL8, MMP9, MMP10, PAI1, SDC1, VEFGA	93	93	Multiplex Protein	Yes	Yes

BTA assays & NMP22 assays

The Bladder tumor antigen (BTA) test is a quantitative ELISA (BTA TRAK) or a qualitative point of care (BTA stat) protein assay that detects urinary complement factor H. Sensitivity reported for BTA is 32-100% with an accompanying specificity of 63-92% [24,25,27-29]. The NMP22 Test and BladderChek test measure urinary NMP22, a nuclear mitotic apparatus protein. The reported sensitivity is 33-100% and specificity is 41-96% [24,25,30-35]. The specificity of these assays may be impacted by other non-cancerous conditions, specifically, hematuria without cancer or inflammation [36-38]. Previous studies have demonstrated a correlation between hematuria and test positivity [39,40]. Rather than detecting a specific bladder cancer biomarker, these assays may be quantifying biomarkers introduced into the urine by a variety of benign conditions, e.g., infection, inflammation, or instrumentation.

UroVysion FISH Assay™

The UroVysion™ Bladder Cancer Kit, is a multicolor, multiprobe fluorescence *in situ* hybridization (FISH) assay that monitors four specific chromosomal alterations in exfoliated urothelial cells. The sensitivity reported for the FISH assay is 51-100% and specificity is 55-100% [24,25,41-49]. Similar with VUC, the lack of sensitivity achieved by UroVysion™ is skewed by the poor detection of low-grade and early stage tumors.

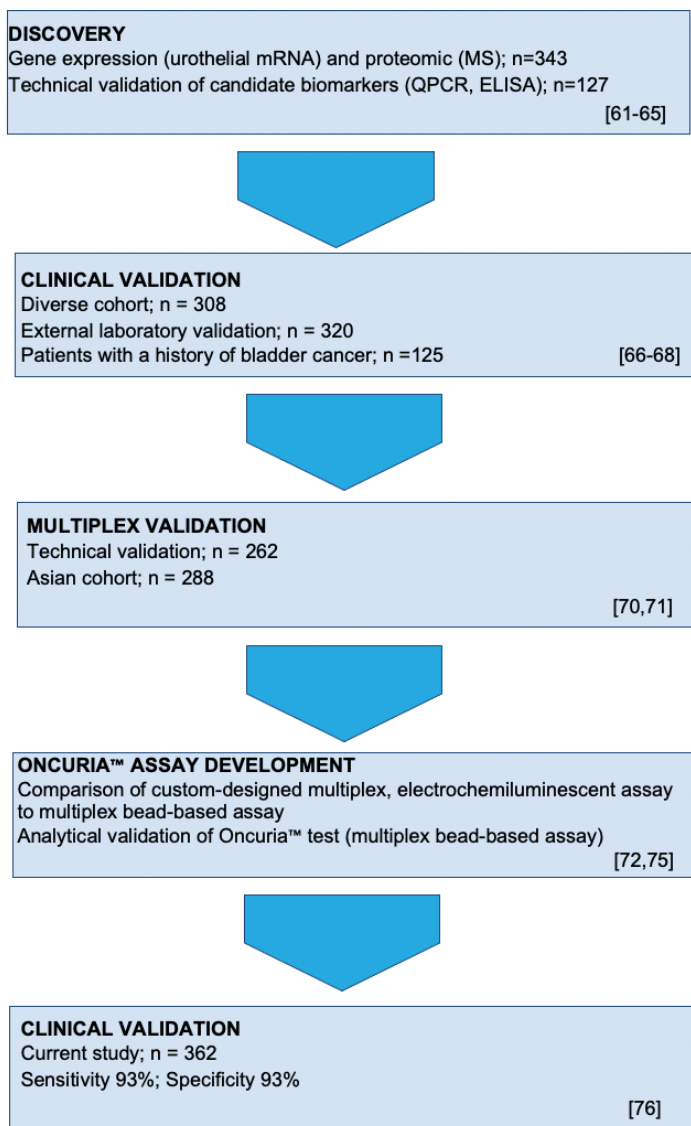
ImmunoCyt Test

The ImmunoCyt/uCyt+ assay is designed to supplement VUC through the detection of cellular biomarkers on cytology slides using fluorescent monoclonal antibodies. The sensitivity reported for ImmunoCyt/uCyt+ is 32-100% with a specificity of 33-91% [24,25,50-53]. As a cell-based assay, ImmunoCyt/uCyt+ is less impacted by hematuria and inflammatory conditions, but, as with cytology, the assay depends on specimen stability and handling, and may be influenced by inter-observer variation.

Multiplex assays

Single biomarkers are limited by the fact that not all bladder cancer tumors, or even those in one category of lesion (e.g., low stage or low-grade) will harbor any single molecular change. Accordingly, the concept that the presence or absence of one molecular biomarker will aid clinical evaluation has not proved to be the case for bladder cancer, or for other cancers. Investigators have begun to identify diagnostic biomarker panels, or

Figure 1. Phased approach for the development and validation of Oncuria™



MS – mass spectrometry; qPCR – quantitative polymerase chain reaction; ELISA – enzyme-linked immunosorbent assay

expression (mRNA) of shed urothelia [61,62], and glycoproteomics profiling of urine supernatant [63,64]. Using sophisticated bioinformatics, the two datasets were combined, and a cancer-associated signature comprised of 19 candidate biomarkers was identified.

The potential clinical utility of the candidate protein biomarkers was monitored in voided urine samples from an independent cohort using commercial ELISA kits. In a cohort of 127 patients (64 with bladder cancer), a 10-protein biomarker signature (ANG; apolipoprotein E, APOE; alpha-1 antitrypsin, A1AT; carbonic anhydrase 9, CA9; interleukin 8, IL8; matrix metalloproteinase 9, MMP9; matrix metalloproteinase 10, MMP10; plasminogen activator inhibitor 1, PAI1; syndecan 1, SDC1 and vascular endothelial growth factor A, VEGFA) achieved a diagnostic sensitivity of 92% and specificity of 97% [65]. Appreciating that benign conditions can adversely affect the performance of urinary biomarkers, signature association was confirmed in a cohort comprised of 108 bladder cancer patients and 202 controls, including patients with urinary tract infection, hematuria and no cancer, kidney stones and moderate to severe voiding symptoms [66], and the potential utility was validated by

signatures. For example, through analysis of nine gene promoters, Hoque *et al.* found that 69% of bladder cancer patients had specific methylation in at least one of four genes; CDKN2A, ARF, MGMT, GSTP1 [54]. By combining the data from all 4-genes, a logistic prediction model was derived that achieved a sensitivity of 82% and a specificity of 96%. Chung *et al.* tested the hypermethylation state of 10 genes in voided urine samples and identified a multigene predictive model comprised of five target genes (*MYO3A*, *CA10*, *NKX6-2*, *DBC1*, and *SOX11*). Sensitivity and specificity of this model were 85% and 95%, respectively [55]. Examples of diagnostic mRNA signatures include those proposed by Hanke *et al.* [56] and Mengual *et al.* [57]. However, these studies have evaluated small and limited populations (*i.e.*, few benign confounding conditions included) and have not undergone extensive validation. Only Holyoake *et al.* have reported extensively on the discovery [58] and validation of a multiplex urinary RNA signature (Cxladder™) to date [59,60].

Derivation of a bladder cancer signature

The methodological approach we deployed to identify a diagnostic bladder cancer signature is depicted in **Figure 1**. Two complementary techniques were applied to profile urine samples from patients with or without bladder cancer; gene

an independent laboratory [67]. The diagnostic signature was also confirmed to perform equally well for the detection of recurrent bladder cancer in a cohort of 125 patients on disease surveillance. The assay outperformed both UroVysion™ and VUC in this context [68]. The analysis of cumulative data from over 1,100 patients confirmed the diagnostic power of the multi-factor signature over individual biomarkers, regardless of histological grade or disease stage of tumors [69], so the development of a robust multiplex assay was initiated. Early prototypes of a multiplex immunoassay were tested in two large independent cohorts. In a US cohort of 200 patients (100 with bladder cancer), the immunoassay achieved a diagnostic sensitivity of 85% and specificity of 81% [70], and in a Japanese cohort of 78 patients (211 with bladder cancer), the next iteration of the immunoassay achieved a diagnostic sensitivity of 85% and specificity of 81% [71]. Furthermore, we have tested and compared the performance of the multiplex assay on alternative technology platforms [72].

Decisions in cancer care depend on certain predictions. Ideally, such predictions or “risks” are personalized, and benefits outweigh the risks in terms of harm or toxicity [73]. Clinical nomograms or risk calculators can provide a probability for certain outcomes, e.g., cancer detection, recurrence, progression, or death. Leveraging the utility of key clinical features (e.g., age, race and gender) in stratifying at-risk patients of harboring bladder cancer, we developed a diagnostic nomogram that was comprised of the 10 biomarker signature plus key clinical features. The addition of molecular data into a clinical nomogram improved the predictive performance, i.e., a hybrid nomogram performed better than demographic or biomarker data alone [74]. Thus, the multiplex immunoassay can be viewed as an In Vitro Diagnostic Multivariate Index Assay (IVDMIA), which combines multiple variables (biomarkers and clinical) using an interpretation function to yield a single, patient-specific result (e.g., a “risk score”), that is intended for use in the diagnosis of disease, in this case bladder cancer.

Oncuria™

The 10-biomarker test has now been developed by BioTechne into a commercial multiplex immunoassay (Oncuria™) for the early detection of bladder cancer in patients presenting with hematuria or with a history of bladder cancer on disease surveillance. The immunoassay is performed using Luminex xMAP technology. The physical components of the BioTechne multiplex assay, a library of detection and capture antibodies, and the secondary reagents have undergone extensive optimization for consistent implementation. Analytical validation of the test has assessed selectivity, sensitivity, specificity, accuracy, linearity, dynamic range, and detection threshold, using voided urine as the test matrix [75]. Lower and upper limits of quantification (LLOQ and ULOQ), antigen cross-reactivity, and the effect of potential interference of the assay by matrix substances has been defined. Clinical validation of the optimized test was performed using urine samples obtained from a cohort of 362 patients (46 with bladder cancer) [76]. In this study, mid-stream voided urine samples were collected from the 362 subjects presenting to outpatient Urology clinics at two sites within the US and were analyzed using the Oncuria™ test. The median age of bladder cancer subjects was 69 years (range 38-87 years), 76.1% were men and 67.4% were Caucasian. Of the 46 bladder cancer cases, 61.4% were classified NMIBC; stages Ta, Tis, T1), and 38.6% were MIBC; stage \geq T2, 19.6% cases were reported as low-grade cancer and 80.4% cases as high-grade. Urinary concentrations of all 10 test analytes were elevated in patients with bladder cancer compared to

Table 2 Mean urinary (\pm SD) concentrations of 10 biomarkers assessed by Oncuria™ in cohort of 362 subjects

Biomarker (pg/mL)	Detectable % of samples	Bladder Cancer N = 46		Non-Cancer Control N = 316		P
		Mean	SD	Mean	SD	
MMP9	64.3	1,237.2	2,191.7	143.0	1,304.3	0.002
IL8	84.4	681.0	1,376.4	90.3	582.5	0.006
VEGFA	88.6	1,003.9	2,743.3	127.8	261.3	0.04
CA9	40.6	8,979	35,518	0.843	2.016	0.09
SDC1	99.3	9,461	6,415	8,707	4,455	0.44
PAI1	71.7	1,169.8	2,803.0	29.8	132.9	0.009
APOE	95.7	16,627	35,895	1,014	2,001	0.005
A1AT	93.2	179,562	236,921	33,742	67,463	0.0001
ANG	81.8	1,800.4	3,170.3	194.7	464.7	0.001
MMP10	57.7	52.79	200.47	4.92	8.88	0.12

those without evidence of bladder cancer (**Table 2**) with statistical significance reached for individual analytes MMP9, IL8, VEGFA, PAI1, APOE, A1AT, and ANG.

A combinatorial analysis of all ten biomarkers achieved a sensitivity of 87% and a specificity of 92%. This was further improved upon when three key clinical factors (e.g., age, race, and gender) were included in the clinical nomogram which achieved a sensitivity of 93%, a specificity of 93% and a negative predictive value of 99%.

Bladder cancer is a common cancer with a high rate of recurrence and progression, and the recurrence phenomenon makes it one of the most prevalent cancers worldwide. The development of a robust non-invasive, urine-based assay for the detection of bladder cancer would clearly have a positive impact on the clinical management of patients with bladder cancer. Oncuria™ is a reliable, validated assay which can provide physicians valuable assistance with the management of patients at risk of harboring bladder cancer. The detection of urinary proteins through multiplex array platforms has the potential to be relatively simple to perform and interpret, and affordable. Currently, four large multicenter, international prospective clinical trials (NCT 03193515, 03193528, 03193541 and NCT04564781) are underway to establish the potential role of Oncuria™ in the physicians' armamentarium.

Reference

1. Wong MCS, Fung FDH, Leung C, Cheung WWL, Goggins WB, Ng CF. The global epidemiology of bladder cancer: a jointpoint regression analysis of its incidence and mortality trends and projection. *Sci Rep*. 2018;8(1):1129. Published 2018 Jan 18. doi:10.1038/s41598-018-19199-z
2. Aben, K. K. and Kiemeny, L. A. Epidemiology of bladder cancer. *Eur Urol*, 36: 660-672, 1999.
3. Letašiová S, Medve'ová A, Šovčíková A, et al. Bladder cancer, a review of the environmental risk factors. *Environ Health*. 2012;11 Suppl 1(Suppl 1):S11. Published 2012 Jun 28. doi:10.1186/1476-069X-11-S1-S11
4. Oțelea MR, Jinga V, Rașcu AȘC, Pleșea IE, Petrescu AN, Mitrache LE, Olteanu M, Bondari D, Rașcu A. Occupational exposure to urinary bladder carcinogens - risk factors, molecular mechanisms and biomarkers. *Rom J Morphol Embryol*. 2018;59(4):1021-1032. Review.
5. Siegel RL, Miller KD, Fuchs HE, Jemal A. Cancer statistics, 2021. *CA Cancer J Clin*. 2021; 71:7-33.
6. Jemal A, Thomas A, Murray T, Thun M. Cancer statistics, 2002. *CA Cancer J Clin*. 2002 Jan-Feb;52(1):23-47. Erratum in: *CA Cancer J Clin* 2002 Mar-Apr;52(2):119. *CA Cancer J Clin* 2002 May-Jun;52(3):181-2.
7. GBoDHFC Network. Evolution and patterns of global health financing 1995-2014: development assistance for health, and government, prepaid private, and out-of-pocket health spending in 184 countries. *Lancet* 2017; 389:1981-2004.
8. "Health Expenditure and Financing" Organisation for Economic Cooperation and Development. <http://stats.oecd.org/Index.aspx?DataSetCode=SHA>. Accessed July 15, 2017.
9. Hall MC, Chang SS, Dalbagni G, Pruthi RS, Seigne JD, Skinner EC, Wolf JS Jr, Schellhammer PF. Guideline for the management of nonmuscle invasive bladder cancer (stages Ta, T1, and Tis): 2007 update. *J Urol*. 2007 Dec;178(6):2314-30.
10. Babjuk M, Burger M, Zigeuner R, Shariat SF, van Rhijn BW, Compérat E, Sylvester RJ, Kaasinen E, Böhle A, Palou Redorta J, Rouprêt M; European Association of Urology. EAU guidelines on non-muscle-invasive urothelial carcinoma of the bladder: update 2013. *Eur Urol*. 2013 Oct;64(4):639-53.
11. Witjes JA. Management of BCG failures in superficial bladder cancer. *Eur Urol* 2006;49:790-7.
12. Zlotta AR, Fleshner NE, Jewett MA. The management of BCG failure in non-muscle-invasive bladder cancer: an update. *Can Urol Assoc J* 2009;3(Suppl4):S199-205
13. Herr HW, Wartinger DD, Fair WR, Oettgen HF. Bacillus Calmette-Guerin therapy for superficial bladder cancer: a 10-year followup. *J Urol*. 1992 Apr;147(4):1020-3.
14. Gupta A, Lotan Y, Bastian PJ, et al. Outcomes of patients with clinical T1 grade 3 urothelial cell bladder carcinoma treated with radical cystectomy. *Urology* 2008;71:302-7.
15. Flaig TW, Spiess PE, Agarwal N, Bangs R, Boorjian SA, Buyyounouski MK, Chang S, Downs TM, Efstathiou JA, Friedlander T, Greenberg RE, Guru KA, Guzzo T, Herr HW, Hoffman-Censits J, Hoimes C, Inman BA, Jimbo M, Kader AK, Lele SM, Michalski J, Montgomery JS, Nandagopal L, Pagliaro LC, Pal SK, Patterson A, Plimack ER, Pohar KS, Preston MA, Sexton WJ, Sieffer-Radtke AO, Tward J, Wright JL, Gurski LA, Johnson-Chilla A. Bladder Cancer, Version 3.2020, NCCN Clinical Practice Guidelines in Oncology. *J Natl Compr Canc Netw*. 2020 Mar;18(3):329-354. doi:10.6004/jnccn.2020.0011.
16. von der Maase H, Sengelov L, Roberts JT, Ricci S, Dogliotti L, Oliver T, Moore MJ, Zimmermann A, Arning M. Long-term survival results of a randomized trial comparing gemcitabine plus cisplatin, with methotrexate, vindesine, doxorubicin, plus cisplatin in patients with bladder cancer. *J Clin Oncol*. 2005 Jul 20;23(21):4602-8.
17. Grivas P, Yu EY. Role of Targeted Therapies in Management of Metastatic Urothelial Cancer in the Era of Immunotherapy. *Curr Treat Options Oncol*. 2019 Jun 28;20(8):67. doi:10.1007/s11864-019-0665-y. Review.
18. Pasin E, Josephson DY, Mitra AP, Cote RJ, Stein JP. Superficial bladder cancer: an update on etiology, molecular development, classification, and natural history. *Reviews in urology*. 2008;10(1):31-43. PubMed PMID: 18470273; PMCID: 2312342.
19. Cancer Genome Atlas Research N. Comprehensive molecular characterization of urothelial bladder carcinoma. *Nature*. 2014;507(7492):315-22. doi:10.1038/nature12965. PubMed PMID: 24476821; PMCID: PMC3962515.
20. PAPANICOLAOU GN. Cytology of the urine sediment in neoplasms of the urinary tract. *J Urol*. 1947 Feb;57(2):375-9. No abstract available.
21. Wiener HG, Vooijs GP, van't Hof-Grootenboer B. Accuracy of urinary cytology in the diagnosis of primary and recurrent bladder cancer. *Acta Cytol*. 1993 Mar-Apr;37(2):163-9.
22. Rife CC, Farrow GM, Utz DC. Urine cytology of transitional cell neoplasms. *Urol Clin North Am*. 1979 Oct;6(3):599-612.
23. Nakamura K, Kasraeian A, Iczkowski KA, Chang M, Pendleton J, Anai S, Rosser CJ. Utility of serial urinary cytology in the initial evaluation of the patient with microscopic hematuria. *BMC Urol*. 2009 Sep 10;9:12. doi:10.1186/1471-2490-9-12.
24. Chou R, Gore JL, Buckley D, Fu R, Gustafson K, Griffin JC, Grusing S, Selph S. Urinary Biomarkers for Diagnosis of Bladder Cancer: A Systematic Review and Meta-analysis. *Ann Intern Med*. 2015 Dec 15;163(12):922-31. doi:10.7326/M15-0997. Epub 2015 Dec 15. PMID: 26501851.
25. Ye F, Wang L, Castillo-Martin M, et al. Biomarkers for bladder cancer management: present and future. *Am J Clin Exp Urol*. 2014;2(1):1-14. Published 2014 Apr 5.
26. Elias K, Svatek RS, Gupta S, Ho R, Lotan Y. High-risk patients with hematuria are not evaluated according to guideline recommendations. *Cancer*. 2010 Jun 15;116(12):2954-9.
27. Thomas L, Leyh H, Marberger M, Bombardieri E, Bassi P, Pagano F, et al. Multicenter trial of the quantitative BTA TRAK assay in the detection of bladder cancer. *Clin Chem*. 1999 Apr;45(4):472-7.
28. Mahner B, Tauber S, Kriegmair M, Nagel D, Holdenrieder S, Hofmann K, et al. Measurements of complement factor H-related protein (BTA-TRAK assay) and nuclear matrix protein (NMP22 assay)—useful diagnostic tools in the diagnosis of urinary bladder cancer? *Clin Chem Lab Med*. 2003 Jan;41(1):104-10.
29. Ellis WJ, Blumenstein BA, Ishak LM, Enfield DL. Clinical evaluation of the BTA TRAK assay and comparison to voided urine cytology and the Bard BTA test in patients with recurrent bladder tumors. The Multi Center Study Group. *Urology*. 1997 Dec;50(6):882-7.
30. Ponsky LE, Sharma S, Pandrangi L, Kedia S, Nelson D, Agarwal A, et al. Screening and monitoring for bladder cancer: refining the use of NMP22. *J Urol*. 2001 Jul;166(1):75-8.
31. Chang YH, Wu CH, Lee YL, Huang PH, Kao YL, Shiao MY. Evaluation of nuclear matrix protein-22 as a clinical diagnostic marker for bladder cancer. *Urology*. 2004 Oct;64(4): 687-92.
32. Berezney R, Coffey DS. Identification of a nuclear protein matrix. *Biochem Biophys Res Commun*. 1974 Oct 23;60(4):1410-7.
33. Grossman HB, Messing E, Soloway M, Tomera K, Katz G, Berger Y, et al. Detection of bladder cancer using a point-of-care proteomic assay. *JAMA*. 2005 Feb 16;293(7):810-6.
34. Grossman HB, Soloway M, Messing E, Katz G, Stein B, Kassabian V, et al. Surveillance for recurrent bladder cancer using a point-of-care proteomic assay. *JAMA*. 2006 Jan 18;295(3):299-305.
35. Shariat SF, Marberger MJ, Lotan Y, Sanchez-Carbayo M, Zippe C, Lüdecke G, Boman H, Sawczuk I, Friedrich MG, Casella R, Mian C, Eissa S, Akaza H, Serretta V, Huland H, Hedelin H, Raina R, Miyanaga N, Sagalowsky AI, Roehrborn CG, Karakiewicz PI. Variability in the performance of nuclear matrix protein 22 for the detection of bladder cancer. *J Urol*. 2006 Sep;176(3):919-26.
36. Atsu N, Ekici S, Oge OO, Ergen A, Hascelik G, Ozen H. False-positive results of the NMP22 test due to hematuria. *J Urol*. 2002 Feb;167(2 Pt 1):555-8.
37. Oge O, Kozaci D, Gemalmaz H. The BTA stat test is nonspecific for

- hematuria: an experimental hematuria model. *J Urol.* 2002 Mar;167(3):1318-9; discussion 9-20.
38. Hennenlotter J, Huber S, Todenhofer T, Kuehs U, Schilling D, Aufderklamm S, et al. Point-of-Care Tests for Bladder Cancer: The Influencing Role of Hematuria. *Adv Urol.* 2011;2011:937561.
 39. Miyake M, Goodison S, Gomes Giacoia E, Rizwani W, Ross S, Rosser CJ. Influencing factors on the NMP-22 urinalysis assay: an experimental model. *BMC Urol.* 2012 Aug 28;12(1):23.
 40. Miyake M, Goodison S, Rizwani W, Ross S, Bart Grossman H, Rosser CJ. Urinary BTA: indicator of bladder cancer or of hematuria. *World J Urol.* 2012 Aug 30.
 41. Junker K, Fritsch T, Hartmann A, Schulze W, Schubert J. Multicolor fluorescence in situ hybridization (M-FISH) on cells from urine for the detection of bladder cancer. *Cytogenet Genome Res.* 2006;114(3-4):279-83.
 42. Caraway NP, Khanna A, Fernandez RL, Payne L, Bassett RL, Jr., Zhang HZ, et al. Fluorescence in situ hybridization for detecting urothelial carcinoma: a clinicopathologic study. *Cancer Cytopathol.* 2010 Jul 21.
 43. Halling KC, King W, Sokolova IA, Meyer RG, Burkhardt HM, Halling AC, et al. A comparison of cytology and fluorescence in situ hybridization for the detection of urothelial carcinoma. *J Urol.* 2000 Nov;164(5):1768-75.
 44. Sarosdy MF, Kahn PR, Ziffer MD, Love WR, Barkin J, Abara EO, et al. Use of a multitarget fluorescence in situ hybridization assay to diagnose bladder cancer in patients with hematuria. *J Urol.* 2006 Jul;176(1):44-7.
 45. Laudadio J, Keane TE, Reeves HM, Savage SJ, Hoda RS, Lage JM, et al. Fluorescence in situ hybridization for detecting transitional cell carcinoma: implications for clinical practice. *BJU Int.* 2005 Dec;96(9):1280-5.
 46. Halling KC, King W, Sokolova IA, Karnes RJ, Meyer RG, Powell EL, et al. A comparison of BTA stat, hemoglobin dipstick, telomerase and UroVysion assays for the detection of urothelial carcinoma in urine. *J Urol.* 2002 May;167(5):2001-6.
 47. Riesz P, Lotz G, Paska C, Szendroi A, Majoros A, Nemeth Z, et al. Detection of bladder cancer from the urine using fluorescence in situ hybridization technique. *Pathol Oncol Res.* 2007;13(3):187-94.
 48. Moonen PM, Merckx GF, Peelen P, Karthaus HF, Smeets DF, Witjes JA. UroVysion compared with cytology and quantitative cytology in the surveillance of non-muscle-invasive bladder cancer. *Eur Urol.* 2007 May;51(5):1275-80; discussion 80.
 49. Fritsche HM, Burger M, Dietmaier W, Denzinger S, Bach E, Otto W, et al. Multicolor FISH (UroVysion) facilitates follow-up of patients with high-grade urothelial carcinoma of the bladder. *Am J Clin Pathol.* 2010 Oct;134(4):597-603.
 50. Greene KL, Berry A, Konety BR. Diagnostic Utility of the ImmunoCyt/uCyt+ Test in Bladder Cancer. *Rev Urol.* 2006 Fall;8(4):190-7.
 51. Li HX, Li M, Li CL, Ma JH, Wang MR, Rao J, et al. ImmunoCyt and cytokeratin 20 immunocytochemistry as adjunct markers for urine cytologic detection of bladder cancer: a prospective study. *Anal Quant CytolHistol.* 2010 Feb;32(1):45-52.
 52. Hautmann S, Toma M, Lorenzo Gomez MF, Friedrich MG, Jaekel T, Michl U, et al. Immunocyt and the HA-HAase urine tests for the detection of bladder cancer: a side-by-side comparison. *Eur Urol.* 2004 Oct;46(4):466-71.
 53. Lodde M, Mian C, Comploj E, Palermo S, Longhi E, Marberger M, et al. uCyt+ test: alternative to cystoscopy for less-invasive follow-up of patients with low risk of urothelial carcinoma. *Urology.* 2006 May;67(5):950-4.
 54. Hoque MO, Begum S, Topaloglu O, Chatterjee A, Rosenbaum E, Van Criekinge W, Westra WH, Schoenberg M, Zahurak M, Goodman SN, Sidransky D. Quantitation of promoter methylation of multiple genes in urine DNA and bladder cancer detection. *J Natl Cancer Inst.* 2006 Jul 19;98(14):996-1004.
 55. Chung W, Bondaruk J, Jelinek J, Lotan Y, Liang S, Czerniak B, Issa JP. Detection of bladder cancer using novel DNA methylation biomarkers in urine sediments. *Cancer Epidemiol Biomarkers Prev.* 2011 Jul;20(7):1483-91.
 56. Hanke M, Kausch I, Dahmen G, Jocham D, Warnecke JM. Detailed technical analysis of urine RNA-based tumor diagnostics reveals ETS2/urokinase plasminogen activator to be a novel marker for bladder cancer. *Clin Chem.* 2007 Dec;53(12):2070-7.
 57. Mengual L, Bursat M, Ribal MJ, Ars E, Marin-Aguilera M, Fernandez M, et al. Gene expression signature in urine for diagnosing and assessing aggressiveness of bladder urothelial carcinoma. *Clin Cancer Res.* 2010 May 1;16(9):2624-33.
 58. Holyoake A, O'Sullivan P, Pollock R, Best T, Watanabe J, Kajita Y, et al. Development of a multiplex RNA urine test for the detection and stratification of transitional cell carcinoma of the bladder. *Clin Cancer Res.* 2008 Feb 1;14(3):742-9.
 59. O'Sullivan P, Sharples K, Dalphin M, Davidson P, Gilling P, Cambridge L, Harvey J, Toro T, Giles N, Luxmanan C, Alves CF, Yoon HS, Hinder V, Masters J, Kennedy-Smith A, Beaven T, Guilford PJ. A multigene urine test for the detection and stratification of bladder cancer in patients presenting with hematuria. *J Urol.* 2012 Sep;188(3):741-7. doi: 10.1016/j.juro.2012.05.003. Epub 2012 Jul 19.
 60. Breen V, Kasabov N, Kamat AM, Jacobson E, Suttie JM, O'Sullivan PJ, Kavalieris L, Darling DG. A holistic comparative analysis of diagnostic tests for urothelial carcinoma: a study of Cxbladder Detect, UroVysion® FISH, NMP22® and cytology based on imputation of multiple datasets. *BMC Med Res Methodol.* 2015 May 12;15:45. doi: 10.1186/s12874-015-0036-8.
 61. Rosser CJ, Liu L, Sun Y, Villicana P, McCullers M, Porvasnik S, Young PR, Parker AS, Goodison S. Bladder Cancer-Associated Gene Expression Signatures Identified by Profiling of Exfoliated Urothelia. *Cancer Epidemiol Biomarkers Prev.* 2009 Feb 3.
 62. Urquidi V, Goodison S, Cai Y, Sun Y, Rosser CJ. A candidate molecular biomarker panel for the detection of bladder cancer. *Cancer Epidemiol Biomarkers Prev.* 2012 Dec;21(12):2149-58.
 63. Kreunin P, Zhao J, Rosser CJ, Urquidi V, Lubman DM, Goodison S. Bladder Cancer Associated Glycoprotein Signatures revealed by Urinary Proteomic Profiling. *J Proteome Res.* 2007 Jul;6(7):2631-9.
 64. Yang N, Feng S, Shedden K, Xie X, Liu Y, Rosser CJ, Lubman DM, Goodison S. Urinary Glycoprotein Biomarker Discovery for Bladder Cancer Detection Using LC/MS-MS and Label-Free Quantification. *Clin Cancer Res.* 2011 May 15;17(10):3349-59.
 65. Goodison S, Chang M, Dai Y, Urquidi V, Rosser CJ. A multi-analyte assay for the non-invasive detection of bladder cancer. *PLoS One.* 2012;7(10):e47469.
 66. Rosser CJ, Ross S, Chang M, Dai Y, Mengual L, Zhang G, Kim J, Urquidi V, Alcaraz A, Goodison S. Multiplex protein signature for the detection of bladder cancer in voided urine samples. *J Urol.* 2013 Dec;190(6):2257-62.
 67. Chen LM, Chang M, Dai Y, Chai KX, Dyrskjot L, Sanchez-Carbayo M, Szarvas T, Zwarthoff EC, Lokeswhar V, Jeronimo C, Parker AS, Ross S, Borre M, Orntoft TF, Jaeger T, Beukers W, Lopez LE, Henrique R, Young PR, Urquidi V, Goodison S, Rosser CJ. External validation of a multiplex urinary protein panel for the detection of bladder cancer in a multicenter cohort. *Cancer Epidemiol Biomarkers Prev.* 2014 Sep;23(9):1804-12.
 68. Rosser CJ, Chang M, Dai Y, Ross S, Mengual L, Alcaraz A, Goodison S. Urinary protein biomarker panel for the detection of recurrent bladder cancer. *Cancer Epidemiol Biomarkers Prev.* 2014 Jul;23(7):1340-5.
 69. Masuda N, Ogawa O, Park M, Liu AY, Goodison S, Dai Y, Kozai L, Furuya H, Lotan Y, Rosser CJ, Kobayashi T. Meta-analysis of a 10-plex urine-based biomarker assay for the detection of bladder cancer. *Oncotarget.* 2018 Jan 3;9(6):7101-7111. doi: 10.18632/oncotarget.23872. eCollection 2018 Jan 23.
 70. Shimizu Y, Furuya H, Bryant Greenwood P, Chan O, Dai Y, Thornquist MD, Goodison S, Rosser CJ. A multiplex immunoassay for the non-invasive detection of bladder cancer. *J Transl Med.* 2016 Jan 30;14:31. doi: 10.1186/s12967-016-0783-2.
 71. Goodison S, Ogawa O, Matsui Y, Kobayashi T, Miyake M, Ohnishi S, Fujimoto K, Dai Y, Shimizu Y, Tsukikawa K, Furuya H, Rosser CJ. A multiplex urinary immunoassay for bladder cancer detection: analysis of a Japanese cohort. *J Transl Med.* 2016 Oct 7;14(1):287. PMID: 27717367
 72. Furuya H, Pagano I, Chee K, Kobayashi T, Wong RS, Lee R, Rosser CJ. Comparison of Commercial ELISA Kits, a Prototype Multiplex Electrochemoluminescent Assay, and a Multiplex Bead-Based Immunoassay for Detecting a Urine-Based Bladder-Cancer-Associated Diagnostic Signature. *Diagnosics (Basel).* 2019 Oct 29;9(4). pii: E166. doi: 10.3390/diagnostics9040166.
 73. Fu AZ, Cantor SB, Kattan MW. Use of nomograms for personalized decision-analytic recommendations. *Med Decis Making.* 2010 Mar-Apr;30(2):267-74.
 74. Huang S, Kou L, Furuya H, Yu C, Goodison S, Kattan MW, Garmire L, Rosser CJ. A Nomogram Derived by Combination of Demographic and Biomarker Data Improves the Noninvasive Evaluation of Patients at Risk for Bladder Cancer. *Cancer Epidemiol Biomarkers Prev.* 2016 Sep;25(9):1361-6. doi: 10.1158/1055-9965.EPI-16-0260. Epub 2016.
 75. Furuya H, Tabula L, Lee R, Kralovec P, Ramsden M, Wong R, Rosser CJ. Analytical validation of ONCURIA™ a multiplex bead-based immunoassay for the non-invasive bladder cancer detection. *Pract Lab Med.* 2020 Nov 13;22:e00189. doi: 10.1016/j.plabm.2020.e00189. PMID: 33294574; PMCID: PMC7691749.
 76. Hirasawa Y, Pagano I, Chen R, Sun Y, Dai Y, Gupta A, Tikhonenkov S, Goodison S, Rosser CJ, Furuya H. Diagnostic performance of Oncuria™, a urinalysis test for bladder cancer. *J Transl Med.* 2021 Apr 6;19(1):141. doi: 10.1186/s12967-021-02796-4. PMID: 33823873; PMCID: PMC802533

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